STUDY ON THE EFFECT OF PROCESSED MILK PROTEIN ON ANGIOTENSING CONVERTING ENZYME ACTIVITY IN RATS

BY LOHIMIWUWYENDRAN ATHITHTHAN



Thesis submitted to the University of Sri Jayewardenepwa for the award of the Degree of Master of Philosophy in Biochemistry of June 2008

DECLARATION BY THE CANDIDATE

The work described in this thesis was carried out by me under the supervision of Prof. Hemantha Peiris (Head of the Department of Biochemistry, Faculty of Medical Sciences, University of Sri Jayewardenepura) and Dr. S.D. Jayeratne (Head of the Department of Medicine, Faculty of Medical Sciences, University of Sri Jayewardenepura) and a report on this has not been submitted in whole or in part to any University or any other Institution for another Degree/Diploma.

Signature of the candidate

03/04/09... Date

DECLARATION BY THE SUPERVISORS

We certify that the above statement made by the candidate is true and that this thesis is suitable for submission to the University for the purpose of evaluation.

Signature of the Supervisor

...<u>06\04.\09</u>.. Date

Signature of the Supervisor

.....0.6.\04\09.

TABLE OF CONTENTS

LIST O	F TABLES	x
LIST OF FIGURES		xii
LIST O	LIST OF PLATES	
ABBRE	ABBREVIATIONS	
ACKNO	OWLEDGEMENTS	xvi
ABSTR	ACT	xix
1	INTRODUCTION	01
1.1	General introduction	01
1.2	Justification	05
1.3	Objectives	07
1.3.1	General objective	07
1.3.2	Specific objectives	07
2	LITERATURE REVIEW	08
2.1	Dairy products	08
2.1.1	Chemical composition of cows milk	08
2.1.1.1	Milk proteins	08
2.1.1.2	Milk fat	09
2.1.1.3	Polypeptides	09
2.1.1.4	Sodium	10
2.1.1.5	Potassium	10
2.1.1.6	Magnesium	11

2.1.1.7	Calcium	11
2.1.2	Determination of protein profile	11
2.1.2.1	Total protein determination	11
2.1.2.2	Determination of whey protein	12
2.2	Hypertension and angiotensin converting enzyme	12
2.2.1	Hypertension	12
2.2.2	ACE and angiotensin	13
2.2.2.1	ACE	13
2.2.2.2	Genetic variants of ACE	13
2.2.2.3	Age and ACE activity	14
2.2.2.4	Kinetics of ACE	14
2.2.2.5	Angiotensin	14
2.2.2.6	Bradykinin	15
2.2.2.7	Renin angiotensin system	15
2.3	Hypertension and antihypertensive peptides on angiotensin	15
	converting enzyme actitivy	
2.3.1	Hypertension and fermented milk	15
2.3.2	ACE activity in other studies	18
2.3.2.1	ACE activity on rats	18
2.3.2.2	Mechanism of inhibition of ACE	18
2.4	Angiotensin converting enzyme inhibitors	19
2 4 1	ACE inhibitors	19

2.4.2	Classification of ACE inhibitors	20
2.4.2.1	Effects of ACE inhibitors	21
2.4.2.2	Application in clinical practice	21
2.5	Hypertension associated diseases	23
2.5.1	Atherosclerosis	23
2.6	Fermented milk and functional food	25
2.6.1	Yoghurt and curd	25
2.6.2	Effect of bacteria	25
2.6.3	Bacteria and their role in chemical composition	26
2.6.4	Starter organism and fermented milk	27
2.6.5	Bacteria and probiotics	28
2.6.6	Probiotics	29
2.6.6.1	Probiotics and health	30
2.6.6.2	Probiotics foods	30
2.6.6.3	Effective bacterial action	31
2.6.6.4	Problems associated with probiotics	32
2.6.7	Functional foods	33
2.6.8	Antibiotics in milk	33
2.7	Changes in cholesterol endothelium and atherosclerosis	34
2.7.1	Obesity	34
2.7.2	Antihypertensive drugs	34
2.7.2	Angiotonsin and cigarette smoke	34

3.	MATERIALS AND METHODS	36
3.1	Materials	36
3.1.1	Water	36
3.1.2	Dairy products	36
3.1.3	Experimental animals	36
3.1.4	Animal feed ingredients	36
3.1.5	Microbial medium and glassware	37
3.1.6	Chemicals	37
3.1.7	Special chemicals and enzymes	37
3.2	Methods	38
3.2.1	Kjeldahl procedure for milk	38
3.2.1.1	Digestion	38
3.2.1.2	Distillation	38
3.2.1.3	Titration	39
3.2.1.4	Calculation	39
3.2.2	Determination of Percentage of total fat in curd	39
3.2.3	Determination of moisture content	40
3.2.4	Separation and isolation of milk proteins	40
3.2.4.1	Preparation of skimmed milk	4(
3.2.4.2	Separation of whole casein	4(
3.2.4.3	Preparation of α-casein	4
2244	Propagation of B-casein	4

3.2.4.5	Preparation of κ-casein	41
	Preparation of whole whey	42
3.2.4.6	Preparation of β-lactoglobulin	42
3.2.4.7		42
3.2.4.8	Preparation of α-lactalbumin	43
3.2.5	Preparation of stock solutions, buffers and gels for electrophoresis	43
3.2.5.1	Resolving and stacking gel solutions	
3.2.5.2	Tris buffer 1.5 M pH 8.8	43
3.2.5.3	Tris buffer 1M pH 6.8	43
3.2.5.4	Sodium dodecyl sulphate	43
3.2.5.5	10% ammoniumpersulfate	44
3.2.5.6	N, N'-tetramethylethylenediamine (TEMED)	44
3.2.5.7	Tris glycine electrophoresis buffer (x 5 concentration)	44
3.2.5.8	Gel loading buffer	44
3.2.5.9	Resolving gels (10%) for tris-glycine SDS polyacrylamide gel	44
	electrophoresis (PAGE)	
3.2.5.10	Stacking gel (5%) for tris glycine SDS PAGE	45
3.2.5.11	Sample preparation	45
3.2.5.12	Molecular weight markers	45
3.2.5.13	Fixing solution	45
3.2.5.14		45
3.2.5.15		46
2 2 5 16		46

3.2.5.17	Determination of approximate molecular weight	4	16
3.2.6	Determination of in-vitro ACE activity	2	17
3.2.6.1	Preparation of HEPES buffer	.2	47
3.2.6.2	Preparation of Hipuryl-histidyl-leucine	4	47
3.2.6.3	Milk	¥	47
3.2.6.4	Curd	i i	47
3.2.6.5	Predigested milk		48
3.2.6.6	Predigested curd		48
3.2.6.7	Preparation of predigested casein and fraction of casein		49
3.2.6.8	Preparation of predigested whey proteins		49
3.2.6.9	Preparation of captopril solution		49
3.2.6.10	Standard curve for ACE activity		50
3.2.6.11	Measurement of ACE inhibitory activity		50
3.2.6.12	Calculation for ACE		51
3.2.7	Test for amino acid peptides and proteins		52
3.2.7.1	Amino acid		52
	Polypeptides		52
3.2.7.2	Proteins		53
3.2.7.3			53
3.2.8	In-vivo experiments		53
3.2.8.1	Experimental animals Preparation of a standard diet for the animal study		53
3.2.8.2	Incorporation of a standard diet for the animal study		54
2283	Incorporation of caselli and curd into standard die		

3.2.8.4	Preparation of predigested casein	55
3.2.8.5	Preparation of predigested curd	55
3.2.8.6	Measuring the moisture content of the diet	55
3.2.8.7	Selection of oral dose	55
3.2.8.8	Collection of blood samples	55
3.2.8.9	Storage of blood samples	56
3.2.8.10	Measurement of parameters	56
3.2.8.11	Animal experiment I	58
3.2.8.12	Animal experiment II	59
3.2.8.13	Animal experiment III	59
3.2.8.14	Statistical analysis	59
3.2.9	Study on microbial aspects	60
3.2.9.1	Selection of curd samples	60
3.2.9.2	Preparation of peptone solution	60
3.2.9.3	Preparation of serial dilution	60
3.2.9.4	Milk agar preparation	60
3.2.9.5	Total colony count	61
3.2.9.6	Preparation of potato dextrose agar (PDA)	62
3.2.9.7	Detection of yeasts and moulds	62
3.2.9.8	MacConkey broth	62
3.2.9.9	Detection of coliform bacteria	6.
2 2 0 10	Propagation of brilliant green blue agar (BGB)	6.

3.2.9.11	Detection of Escherichia coli	63
3.2.9.12	Preparation of Indole Reagent (Kovasc)	64
3.2.9.13	Confirmation of the presence of <i>E.coli</i>	64
4.	RESULTS	65
4.1	Protein profiles in milk and curd	65
4.1.1	Composition of milk and curd	65
4.1.2	SDS PAGE and densitogram of milk proteins	65
4.1.3	SDS PAGE and densitogram of curd and curd proteins	74
4.1.4	SDS-PAGE obtained from different brands of curd	82
4.2	In-vitro assay and standard curve	84
4.2.1	Standard curve for ACE activity	84
4.2.2	In-vitro assay of different proteins and their ACE inhibiting activity	84
4.2.3	Presence of amino acid, polypeptides and proteins	85
4.2.4	Standard curves for cholesterol and triglycerides	85
4.3	The in-vivo study on ACE activity, lipid profile, body weight and	87
	feed intake in Wistar rats	
4.3.1	Experiment I-Effect of feeding casein or curd incorporated pellets on	87
****	blood parameters and body weight of rats	
4.3.2	Experiment II-Effect of oral feeding hydrolyzed casein or curd on	88
	Wistar rats	ħ
4.3.3	Experiment III- Effect of oral feeding whey on Wistar rats	98
4.4	Study on microbiological aspects	10

4.4.1	Analysis of total colony count in curd	101
4.4.2	Determination of shelf life of the curd samples	101
4.4.2.1	Results of yeast and mould study	101
4.4.2.2	Results of coliform bacteria and E.coli study	101
5	DISCUSSION	106
6	CONCLUSIONS	118
7	REFFRENCES	121
8	APPENDICES	134

LIST OF TABLES

Table 4.1	Major constituents of milk and curd and pH range on the production	67
27	date and expiry date	
Table 4.2	The in-vitro ACE inhibiting activity of different proteins and curd.	86
Table 4.3	Effect of feeding casein or curd incorporated pellets on body weight,	90
	serum ACE level and lipid profile on Wistar rats at four and eight	
	weeks of experiment I	
Table 4.4	Mean changes in body weight, serum ACE level and lipid profile on	91
	Wistar rats at eight weeks of experiment I	
Table 4.5	Effect of oral feeding of hydrolyzed casein or curd on body weight	94
	serum ACE level and lipid profile on Wistar rats at four and eight	
	weeks of experiment II	
Table 4.6	Mean changes in body weight, serum ACE level and lipid profile on	95
	Wistar rats at eight weeks of experiment II	
Table 4.7	The effect of whey on body weight, serum ACE level and lipid	99
	profile on Wistar rats at eight weeks of experiment III	
Table 4.8	Mean changes in body weight, serum ACE level and lipid profile on	100
	Wistar rats at eight weeks of experiment III	
Table 4.9		103
	production	
Table 4.10	Presence of Yeast and mould in curd on day 0, 7, 14 from date of	104
	production	

Table 4.11 Presence of Coliform bacteria and *E.coli* on day 0,7 and 14 from date 105 of production

LIST OF FIGURES

Figure 1.1	Schematic diagram of RAS and Kallikrein-Kinin system	2
Figure 2.1	The potential activation and inactivation of ACE-I inhibitory peptides	17
	in the human body during gastrointestinal digestion and absorption,	
	and in the blood	
Figure 4.1	Scanned SDS-PAGE of different milk proteins with the molecular	68
	weight markers	
Figure 4.2	Densitogram of α -casein fraction of milk (lane 1 of figure 4.1)	69
Figure 4.3	Densitogram of β -casein fraction of milk (lane 2 of figure 4.1)	70
Figure 4.4	Densitogram of κ -casein fraction of milk (lane 3 of figure 4.1)	71
Figure 4.5	Densitogram of whey fraction of milk (lane 4 of figure 4.1)	72
Figure4.6	Densitogram of molecular weight markers (Lane 5 of figure 4.1)	73
Figure 4.7	Scanned picture of SDS-PAGE of curd, proteins of curd and the	76
	corresponding molecular weight marker	
Figure 4.8	Densitogram of the molecular weight markers lane 6 of figure 4.7	77
Figure 4.9	Densitogram of β -casein fraction of curd, lane 7 of figure 4.7	78
Figure 4.10	Densitogram of fresh curd, lane 8 of figure 4.7	79
Figure 4.11	Densitogram of α -casein fraction of curd, lane 9 of figure 4.7	80
Figure 4.12	Densitogram of κ-casein fraction of curd, lane 10 of figure 4.7	81
Figure 4.13	The basal & end values of serum ACE level of test and control	92
	groups of experiment I	

LIST OF FIGURES

Figure 4.14	The basal & end values of serum cholesterol of test and control	93
	groups of experiment I	
Figure 4.15	The basal & end values of ACE level of test and control groups of	96
	experiment II	
Figure 4.16	The basal & end values of serum cholesterol of test and control	97
	groups of experiment II	

LIST OF PLATES

Plate 4.1	SDS PAGE obtained from different curd brands	82
Plate 4.2	SDS PAGE obtained from casein of different curd brands	83

ABBREVIATION

ACE Angiotensin Converting Enzyme

ANG Angiotensin

APC Aerobic Plate Count

AT Angiotensin receptor

BGB Brilliant green blue

BW Body weight

°C Degrees centigrade

CN Casein

Con Concentrated

CPPs Caseinophosphopeptides

DBP Diastolic Blood Pressure

E.Coli Escherichia Coli

FAO Food and Agriculture Organization

FAPGG Furylacrylolyl-phenylalanyl-glycyl-glycine

GALT Gut Associated Lymphoid Tissue

GFR Glomerular Filtration Rate

GIT Gastrointestinal tract

HDL High Density Lipoproteins

HHL Hippuryl -L-Histidyl-Leucine

HT Hypertension

IL Interleukines

INF Interferon

ABBREVIATION

Ig Immunoglobulin

L Lactobacillus

LA Laboratory Accident

LAB Lactic Acid Bacteria

LDL Low Density Lipoprotein

MA Milk Agar

MI Myocardial Infarction

MNNG N-methyl-N'-nitro-N-nitrosoguanidine

MRI Medical Research Institute

mw Molecular Weight

PAGE Poly Acrylamide Gel Electrophoresis

PDA Potato Dextrose Agar

RAS Renin Angiotensin System

SBP Systolic Blood Pressure

SD Standard Deviation

SDS Sodium Dodecyl Sulphate

SHR Spontaneously Hypertensive Rats

Subsp Sub species

TC Total Cholesterol

TG Triacylglycerol/Triglycerides

TNF Tumour Necrosis Factor

WHO World Health Organization

ACKNOWLEDGEMENT

Firstly I am indebted to my supervisor Prof. Hemantha Peiris (Head of the Department of Biochemistry, Faculty of Medical Sciences, University of Sri Jayewardenepura) for the excellent supervision given to me throughout the project. He generously gave time, patient guidance and wise advice that were never short of being on target.

I wish to express my gratitude to my supervisor Dr. S. D. Jayaratne (Head of the Department of Medicine, Faculty of Medical Sciences, University of Sri Jayewardenepura) who helped me to start the research and gave me necessary advice on clinical aspects and on modification of the protocols at needy times. He generously helped me throughout my research especially in the writing up this thesis and without him this work would not have been feasible.

I sincerely thank Dr. S. Jayesekara (Head of the Animal Centre, Medical Research Institute) for her excellent supervision and training given to me to carry out the animal study successfully. I wish to thank her for giving me the opportunity to utilize the facilities available at their institute. Her encouragement and kind co-operation given to me is highly appreciated.

I would like to sincerely thank the Chairman and Head of the Microbiology Unit of Milco Private Limited, Sri Lanka, for providing me with free bench facilities and supervision at the Microbiology unit which enable me to carry out the microbiology study.

I wish to thank the Director of MRI for granting permission to carry out the study and the staff members of the Animal Centre of MRI, for their support and kind corporation. The Head and the staff of the Department of Microbiology, Faculty of Medical Sciences, University of Sri Jayewardenepura, for providing me with the laboratory facilities to carry

out the electrophoresis work, Head and staff of the Department of Food Science & Technology, Faculty of Applied Sciences, University of Sri Jayewardenepura for providing me with the laboratory facilities to use the Kjeldahl apparatus and Head and the staff of Department of Botany, Faculty of Applied Sciences, University of Sri Jayewardenepura for granting permission to use the centrifuge machine.

I wish to sincerely thank all my research colleagues of Department of Biochemistry, Faculty of Medical Sciences, University of Sri Jayewardenepura, for their support and for sharing their valuable time and knowledge at critical stages.

I wish to thank all the staff members of the Department of Biochemistry specially Prof. E.R.Jansz for their contributions at needed time.

I wish to acknowledge the financial assistance given by the University of Sri Jayewardenenpura (University Grant ASP/06/Re/2004) and the National Science Foundation scholarship (NSF/Sch/2003/04) to carry out the animal study.

Finally I would like to thank my husband and parents for their support, encouragement and for the special care given to my daughter throughout the study. I wish to thank my brothers for their enthusiasm shown in getting some valuable references. With great love I wish to thank my daughter for her tolerance and patience and I dedicate this study to my family members.

STUDY ON THE EFFECT OF PROCESSED MILK PROTEIN ON ANGIOTENSIN CONVERTING ENZYME ACTIVITY IN RATS LOHINI VIJAYENDRAN ATHITHTHAN

ABSTRACT

Milk peptides are known to have antihypertensive effects by inhibiting the Angiotensin Converting Enzyme (ACE) and thereby inhibiting the formation of angiotensin II (ANG II). Due to lack of available literature pertaining to SAARC countries including Sri Lanka on antihypertensive effects of milk proteins, this study was carried out to investigate the effects of milk proteins and curd on ACE activity and their effects on lipid profile using *Wistar* rats.

Milk and curd of a commercially available local brand was used in all the experiments. Protein fractions of milk and curd were isolated and separated using SDS PAGE electrophoresis. Results revealed that the ACE inhibiting activity due to in-vitro digested products, curd, milk, whole casein, whole whey, α-casein, β-casein and κ-casein were 78.98%, 48.09%, 83.6%, 53.5%, 43.2%, 82.1% and 80.1% respectively. Prior to enzymatic digestion, ACE inhibiting activity of milk and curd were 3.99% and 49.41% respectively. The enzymatic digest of total casein which had the highest ACE inhibition (83.6%) in the *in-vitro* assay was further subjected to an animal study. Three different animal experiments were carried out to determine the effects of casein, whey and curd on serum ACE activity and long term intake on the lipid profile of *Wistar* rats. In experiment I, the test groups were fed with World Health Organization (WHO) standard feed incorporated with casein or curd, while the control group received the standard feed. In experiment II, test groups were

orally fed with either 2 ml of hydrolyzed casein or curd whilst experiment III test group received 2 ml of whey while control groups received 2 ml of water in addition to the standard feed. The mean differences obtained for ACE activity for individual animals were analyzed after eight weeks. In Experiment I, casein fed group had a higher serum ACE reduction when compared with the control group. The mean difference of ACE (U/L) in both casein (-37.2 ±33.5) and curd (-12.3±16.2) treated groups were significantly lower (p<0.05) when compared to the control group (54.7±66.2). In experiment I, curd treated group also had a lesser increase in serum cholesterol (1.65 ±12.8g/dL) when compared to the control group (16.5±4.8g/dL) whilst the whey treated group in experiment III had a mean reduction of -2.8±4.0 g/dL for serum total cholesterol. The mean difference in both curd and whey fed groups showed a significantly lower value for serum cholesterol (p<0.05). However, high density lipoprotein cholesterol, triglycerides, feed intake and body weight did not show a significant difference between the animals in the treatment group and the control groups. These results suggest that both casein and curd have an inhibitory effect on serum ACE activity. In addition to the ACE inhibitory effect, curd also has a serum cholesterol lowering effect on Wistar rats.