

A BETTER EARTH IS POSSIBLE

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सत्यमेव जयते



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leaves (6.66 and 8.00) at 30 and 60 DAS respectively. The root length (14.30 cm), fresh and dry root weight (0.55 and 0.20 g) and reduced nematode parameters minimum number of galls per root system (17.33), nematode population and gall index (696.33 and 3.0) at 60 DAS respectively. The combination of treatments with neem cake + carbofuran 3G proved best followed by a bio-control agent and recorded the highest increased growth parameters and reduced nematode parameters. This study shows that organic green manure amendments and bio-control agents will help to reduce nematode population and aid in sustainable crop production.

Microbial control of toxin producing cyanobacterium *Microcystis aeruginosa* in a hypereutrophic lake

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Abstract

The dynamics of cyanophages and algicidal bacteria which infect the bloom forming cyanobacterium *Microcystis aeruginosa* were followed in the Beire Lake (a hypereutrophic lake) from September 2008 to August 2009. Densities of *M. aeruginosa* range between 4.0×10^5 and 1.9×10^7 cells ml⁻¹, while those of algicidal bacteria were between 4.0 and 5.1×10^2 PFU ml⁻¹ and of cyanophages between $< 10^2$ PFU ml⁻¹ and 7.1×10^3 PFU ml⁻¹. A significant relationship was found between the density of algicidal bacteria and of *M. aeruginosa* ($r = 0.81$, $n = 69$, $p < 0.001$) suggesting that the dynamics of algicidal bacteria may regulate the abundance of *M. aeruginosa*. Occasional peaks of cyanophages densities were detected in October, June and August when sharp declines in *M. aeruginosa* cell densities were observed. Densities of cyanophages became undetectable when the abundance of *M. aeruginosa* was low, suggesting the density dependent infection of *M. aeruginosa* by cyanophages. Thus, the present study suggests that infections of both algicidal bacteria and cyanophages are important biological agents which decompose a bloom of *M. aeruginosa* in freshwater environment.

A freshwater gliding bacterium, *Alcaligenes denitrificans*, was isolated from the Beire lake during the study period. This bacterium caused cell lysis and death of some cyanobacterial species, but showed no algicidal effects on the species of chlorophyceae tested. *M. aeruginosa*, *M. viridis* and *M. wesenbergii* were susceptible to the bacterial attack and the growth-inhibiting effect of the bacterium was significant on *M. aeruginosa*, particularly when the alga was in the exponential growth phase. When *A. denitrificans* was inoculated at low densities (10^3 cells ml⁻¹) together with *Microcystis* species, the bacterium proliferated to 10^8 cells ml⁻¹ and caused algal cell lysis. *M. aeruginosa* died when *A. denitrificans* was added to the algal culture but not when only the filtrate from the bacterial culture was added. This suggests that extracellular products are not inhibitory to *M. aeruginosa* and that only direct contact between *A. denitrificans* and *M. aeruginosa* was lethal. Thus, the study suggests that *A. denitrificans* plays an important role influencing the growth of *Microcystis* and contributes to the death of *Microcystis* in freshwater environments.

The effect of microflagellates on the decay of viruses was studied *in vitro* using fractionated natural water. We found considerable decay of VLPs in the 5.0 μm filtrate relative to the 0.2 to 0.8 μm filtrate.